

Method Development and Scale-up for Peptide Purification

Case Studies for a GLP-1 Agonist and a Peptide Hormone

**25.02.
2025**

**11-12 am
(CET)**

Speakers

Julia Bartmann, YMC Europe GmbH

Dr. Yannick Krauke, KNAUER Wissenschaftliche Geräte GmbH

Great Potential of Peptides

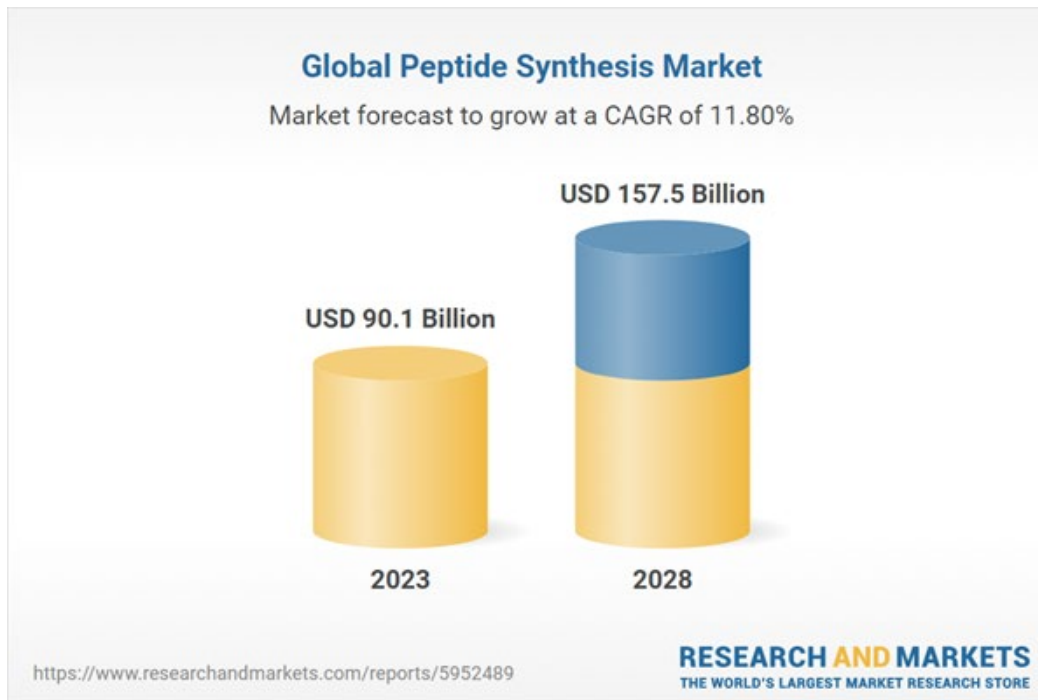


Advantages of peptides as therapeutic agent

- highly specific
- well tolerated
- biodegradable



- cancer therapies
- diabetes management
- neurological disorders
- cardiovascular diseases
- rare genetic conditions



Continuous Chromatography for Liraglutide

BACHEM

PRESENTATION

**A Second-generation Process
for the Manufacturing
of Liraglutide**



Alexander Kleinsmann, PhD
Director R&D



BACHEM

YMC
ChromaCon

at TIDES 2024:

Liraglutide presentation available as
webinar:

<https://www.bachem.com/webinar/peptides-nce/a-second-generation-process-for-the-manufacturing-of-liraglutide/>

Preparative Method Development

1. Method development
at analytical scale



Analytical method needed:

- As basis for preparative process development
- For analysis of collected fractions

2. Loadability studies at
analytical scale



3. Linear scale-up to
preparative process

- Stationary phase selection
- Pore size requirements
- Selection of particle size
- Mobile phase composition
- Gradient optimisation

Preparative Method Development

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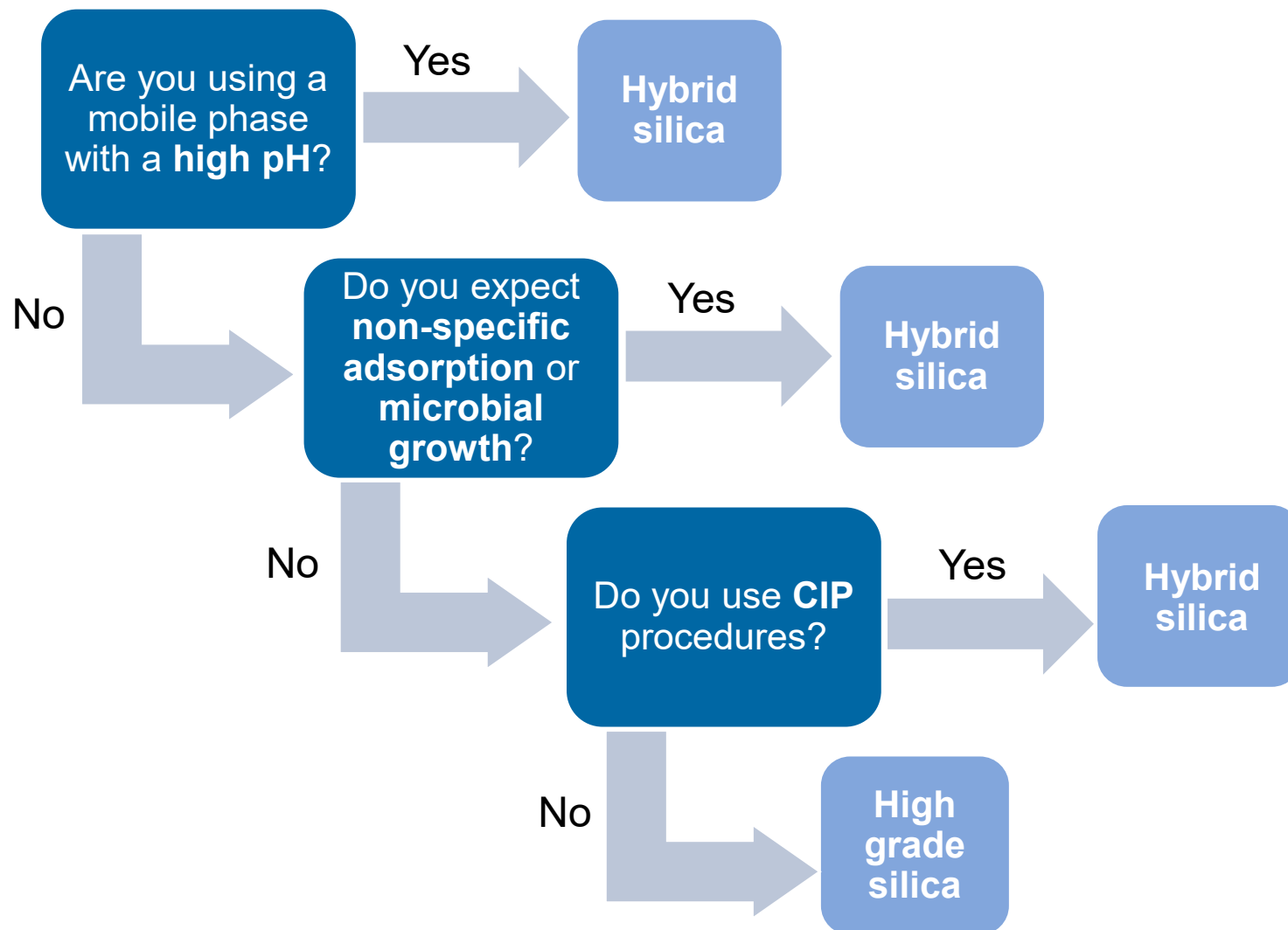


3. Linear scale-up to
preparative process

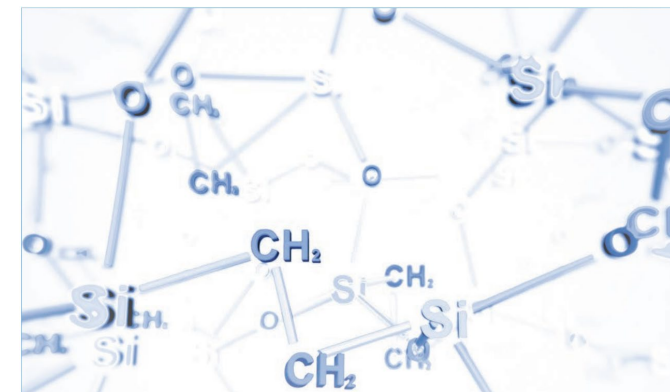
- Stationary phase selection
- Pore size requirements
- Selection of particle size
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- Gradient optimisation

The Choice of the Base Particle

Which considerations should be made?

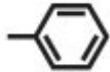






Hybrid silica



organic ethylen bridges
between the silica

Stationary Phase Selection

	C18	C8	Phenyl	C4
Functional group	$-C_{18}H_{37}$	$-C_8H_{17}$		$-C_4H_9$
Hydrophobicity	High			Low
Hydrogen bonding capacity	Low			High
Surface recognition ability	High			Low
Suitable MW of the peptide	Low			High

Most commonly used selectivities for peptides

For peptides with aromatic systems

For very large peptides

→ Screening of different selectivities

Find the Best Selectivity by Column Screening

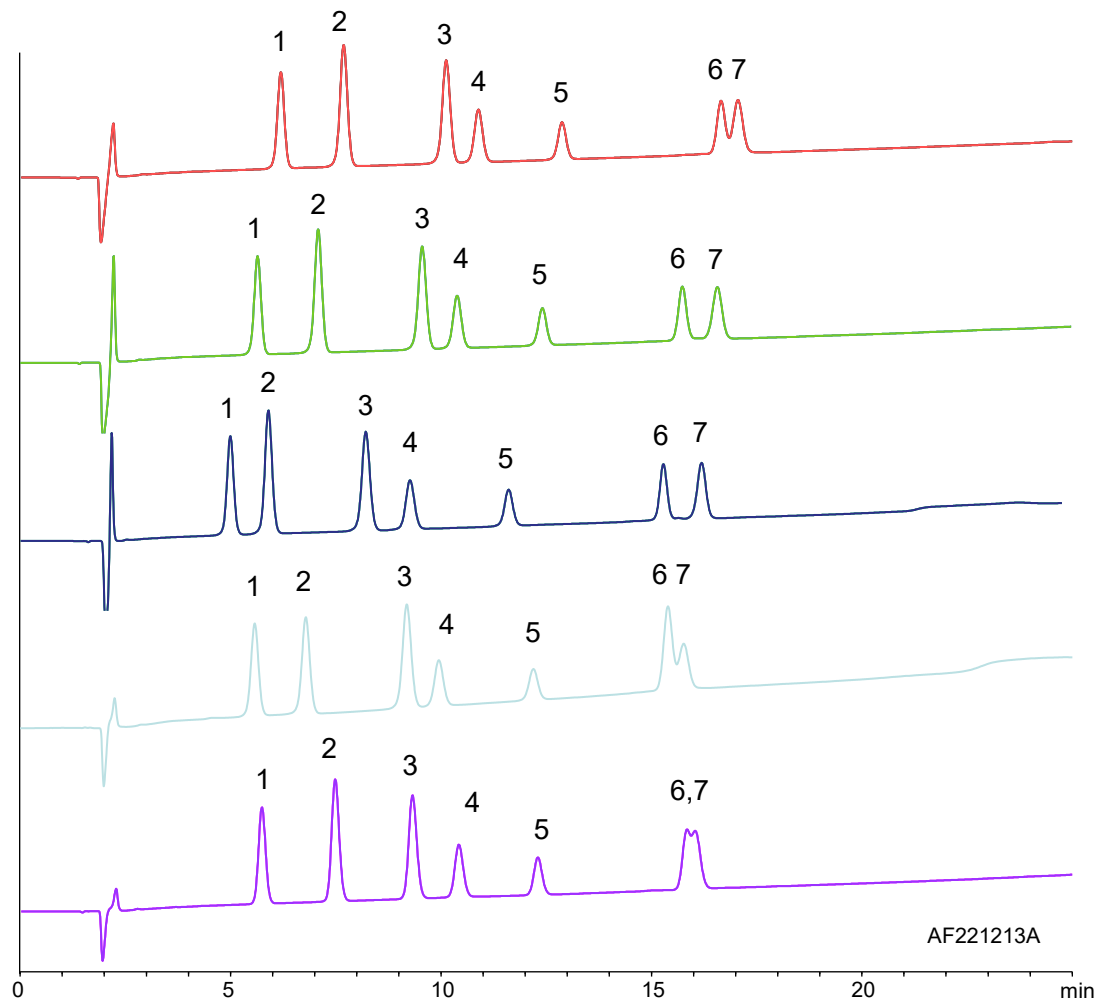
YMC-Triart Prep C18-S

YMC-Triart Prep C8-S

YMC-Triart Prep Bio200 C8

YMC-Triart Prep C4-S

YMC-Triart Prep Phenyl-S



Peptide Sample

- | | |
|------------------------|------------|
| 1. Oxytocin | (MW 1,007) |
| 2. Met-Enkephalin | (MW 574) |
| 3. Leu-Enkephalin | (MW 556) |
| 4. Neurotensin | (MW 1,673) |
| 5. γ -Endorphin | (MW 1,859) |
| 6. Insulin (Bovine) | (MW 5,733) |
| 7. β -Endorphin | (MW 3,465) |

Optimal Selectivity: Aromatic Peptides

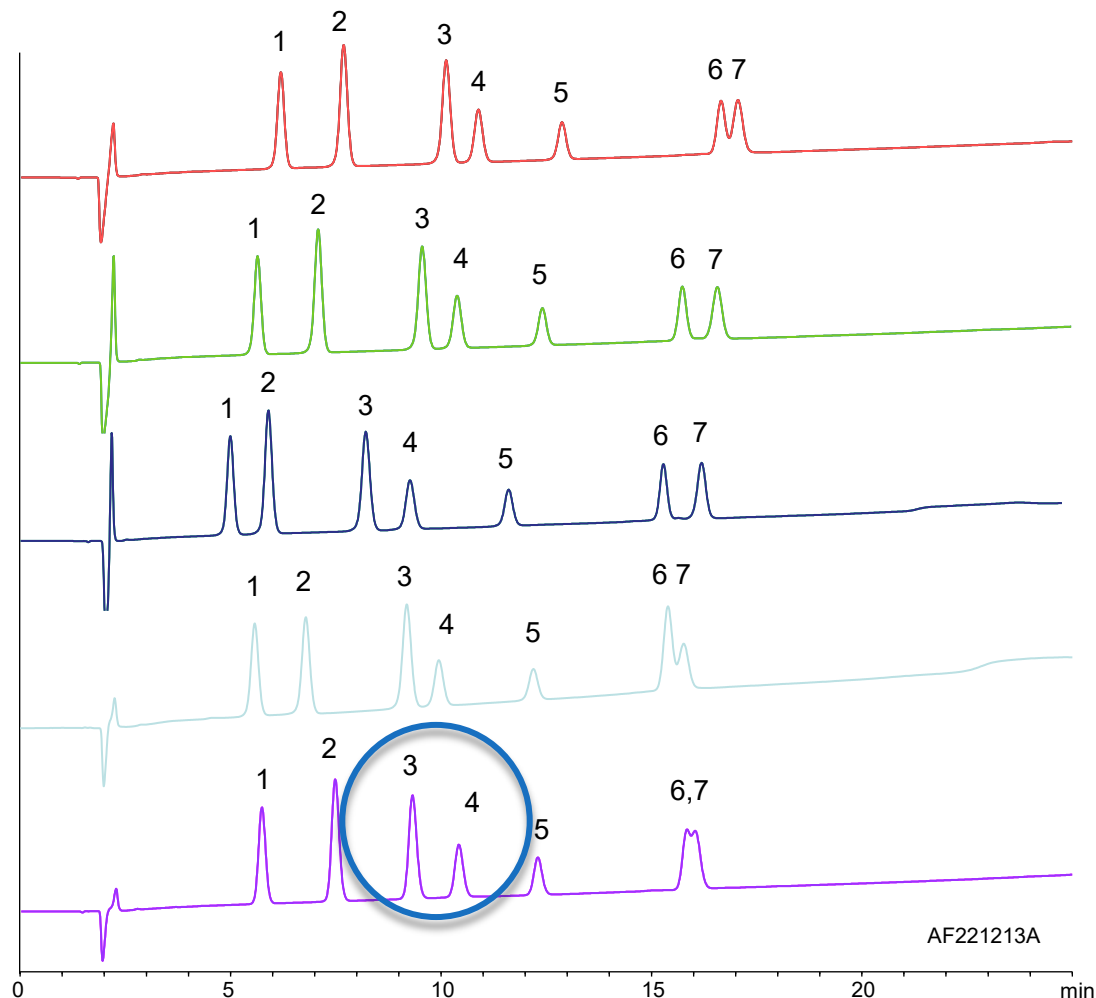
YMC-Triart Prep C18-S

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→ Phenyl modification for π - π -interactions of aromatic groups

Optimal Selectivity: Large Peptides

YMC-Triart Prep C18-S

120 Å

YMC-Triart Prep C8-S

120 Å

YMC-Triart Prep Bio200 C8

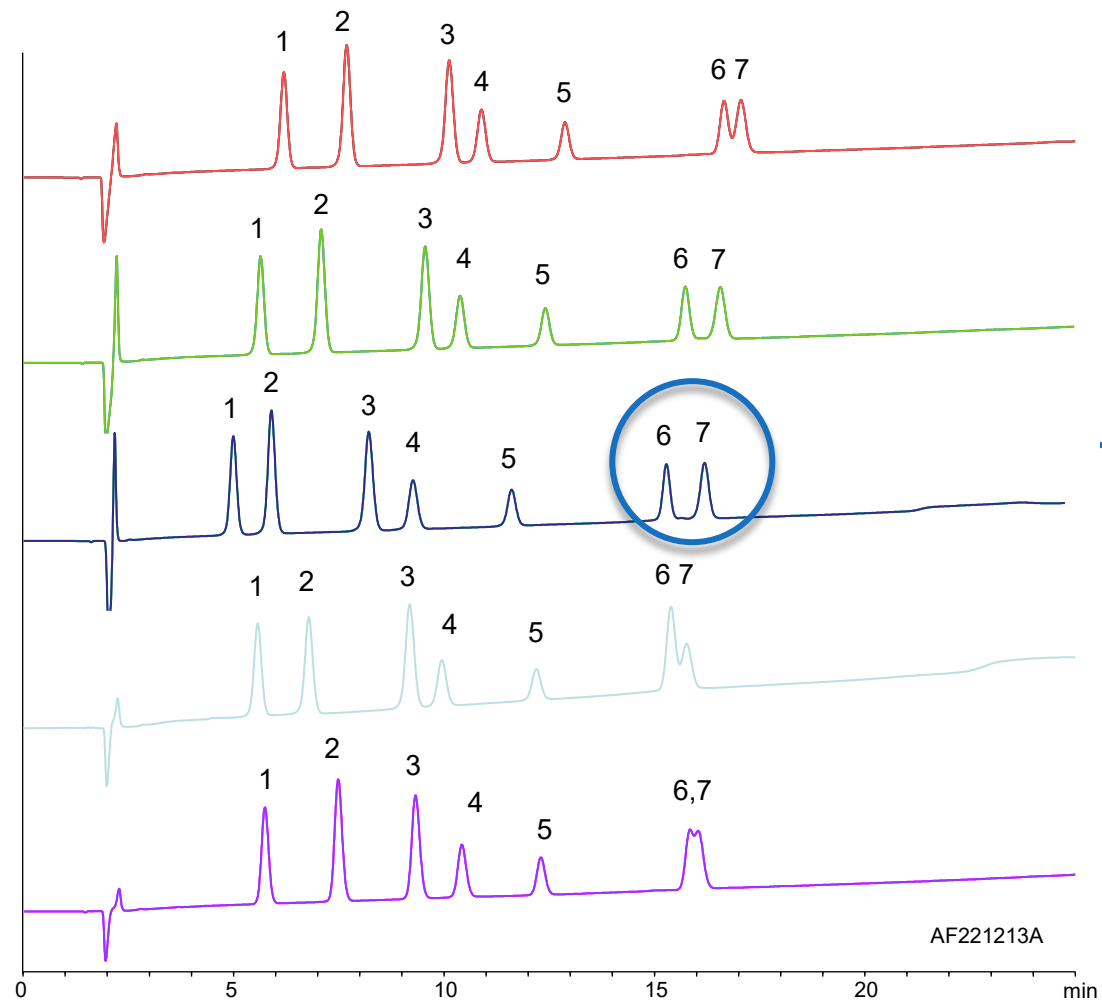
200 Å

YMC-Triart Prep C4-S

120 Å

YMC-Triart Prep Phenyl-S

120 Å



Peptide Sample

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Larger pore size needed!

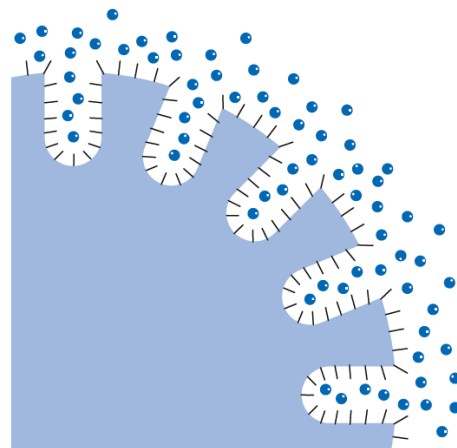
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Selection of Pore Size

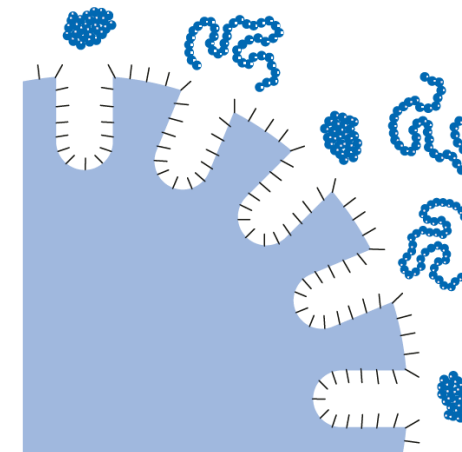
Peptide Size

Consider steric exclusion of the compound from pores and densely modified stationary phase surfaces

Small peptides



Larger peptides and polymers



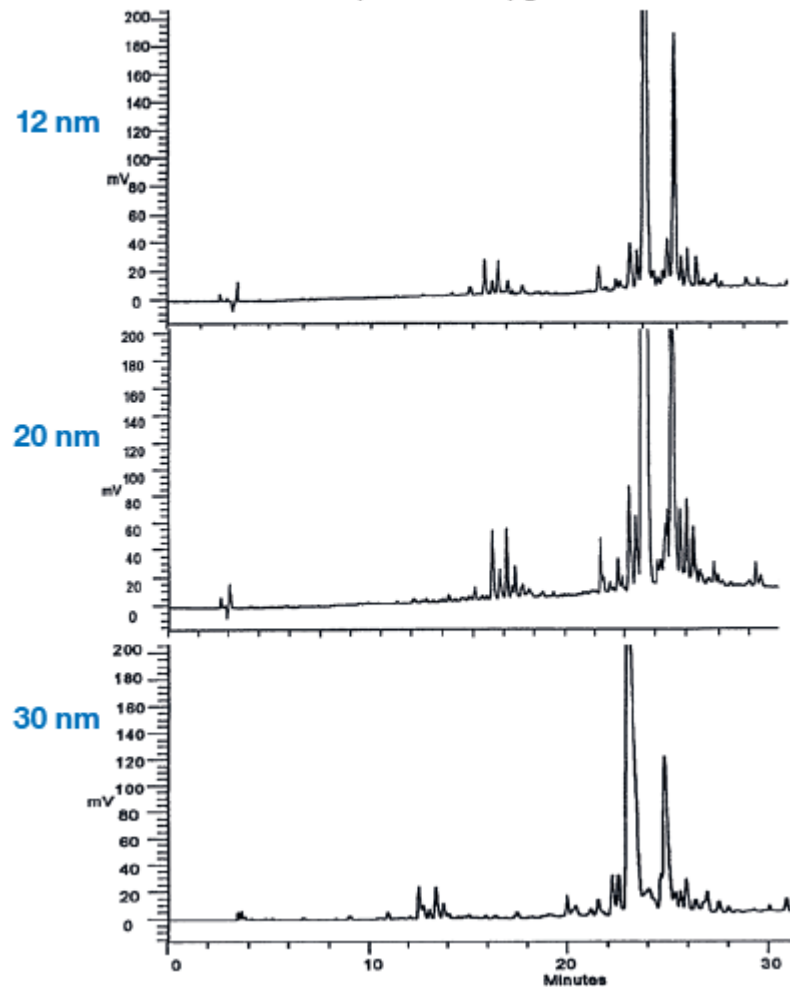
MW			C18	C8	C4
 5,000 20,000 100,000*	12 nm		+++	++	+
	20 nm		++	+++	++
	30 nm		+	++	+++

The pore size of the stationary phase should be selected as large as necessary and as small as possible.

Influence of the Pore Size on Loadability

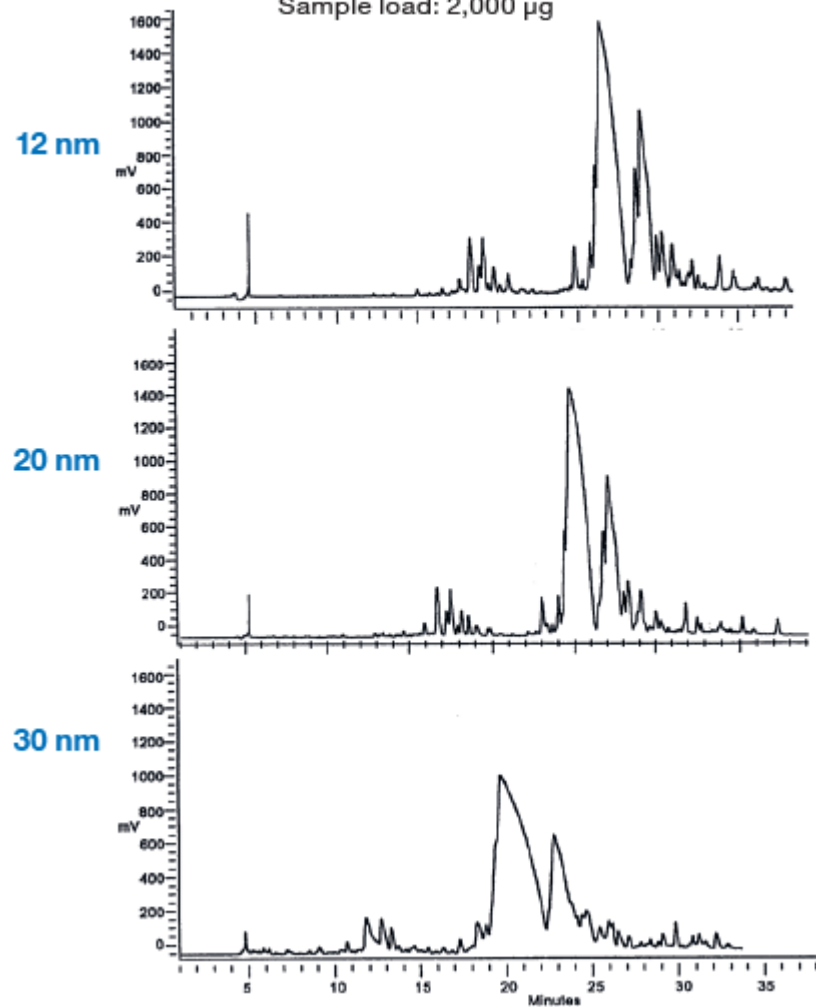
Analytical separation of a peptide

(17 AA, MW 3,400)
Sample load: 50 µg



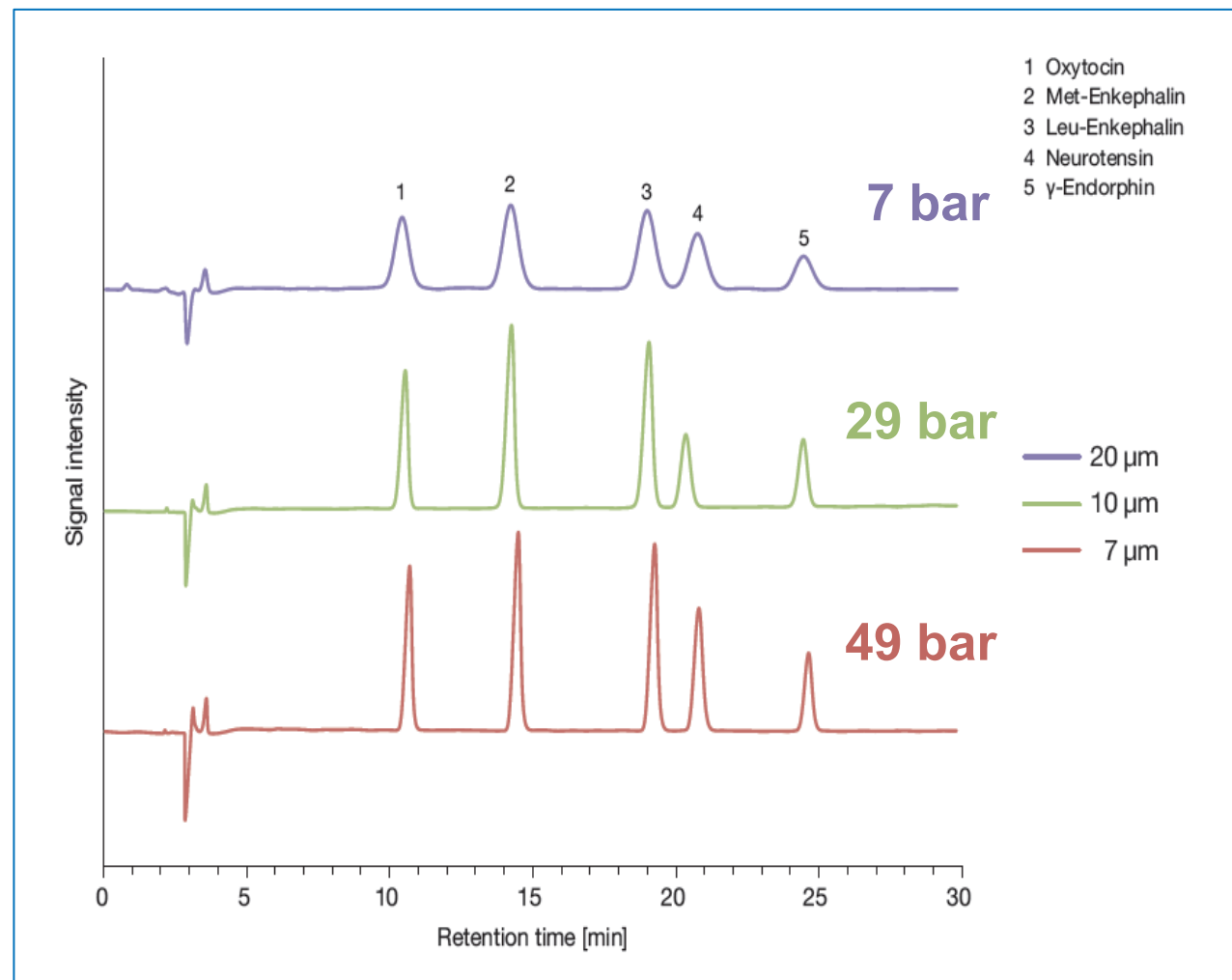
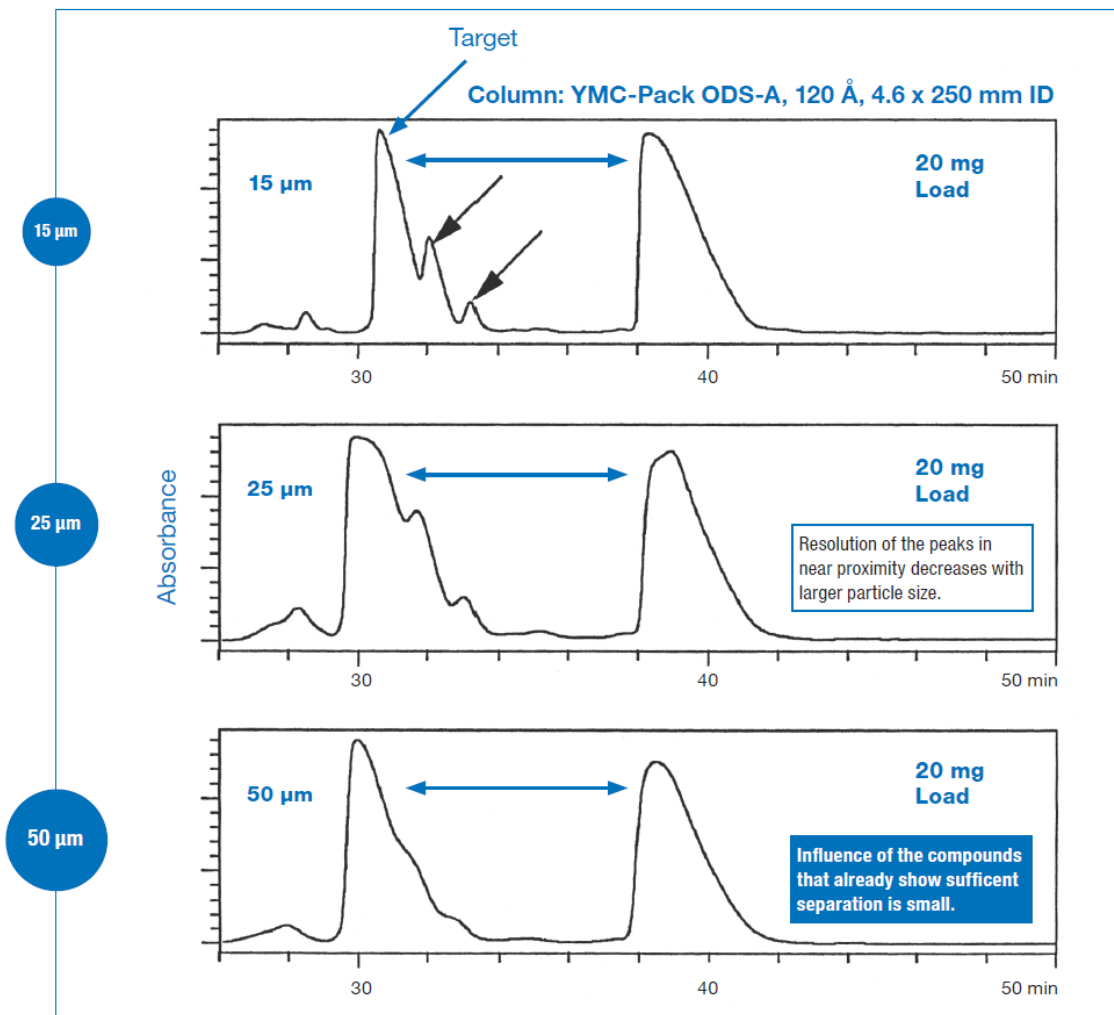
Preparative separation of a peptide

(17 AA, MW 3,400)
Sample load: 2,000 µg



Overloading
Effects!

High Resolution Separations: Optimal Particle Size



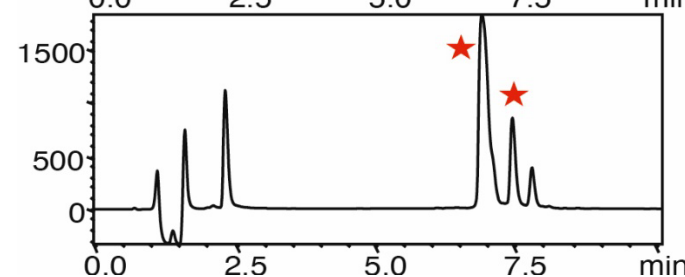
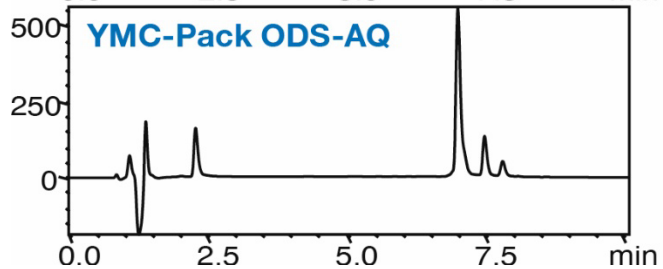
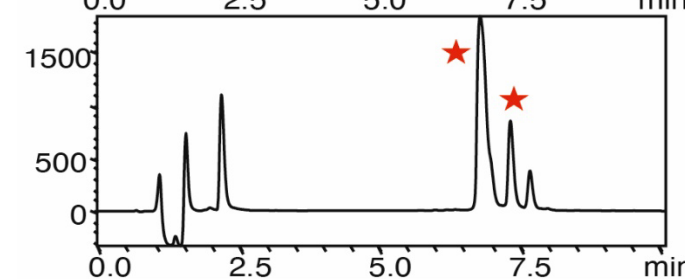
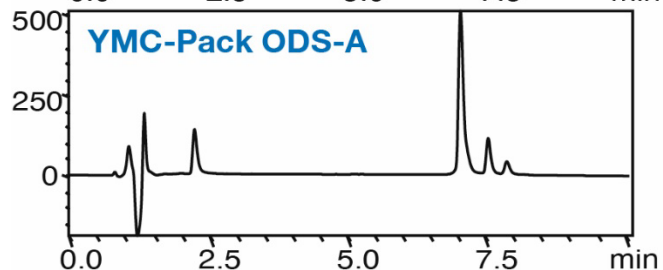
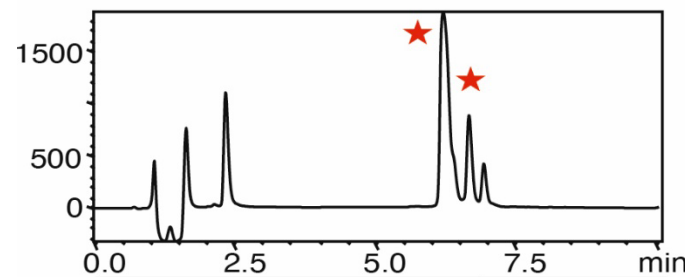
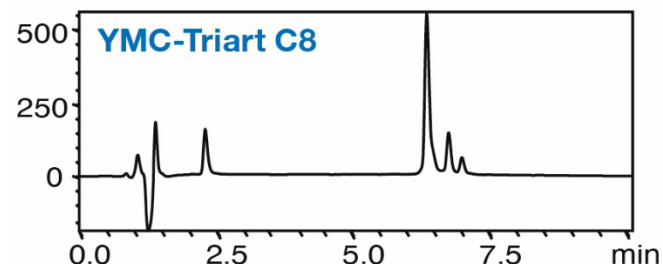
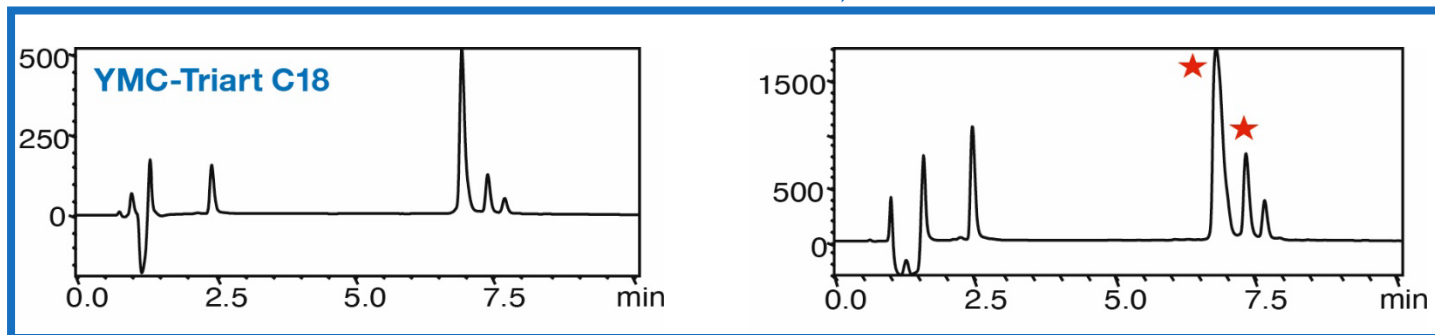
Phase Screening for Liraglutide

Loading amount

3 μ g

x 8 Loading

24 μ g



Not only the resolution is important, but also the pH stability!



Higher flexibility in method development

★ critical pair

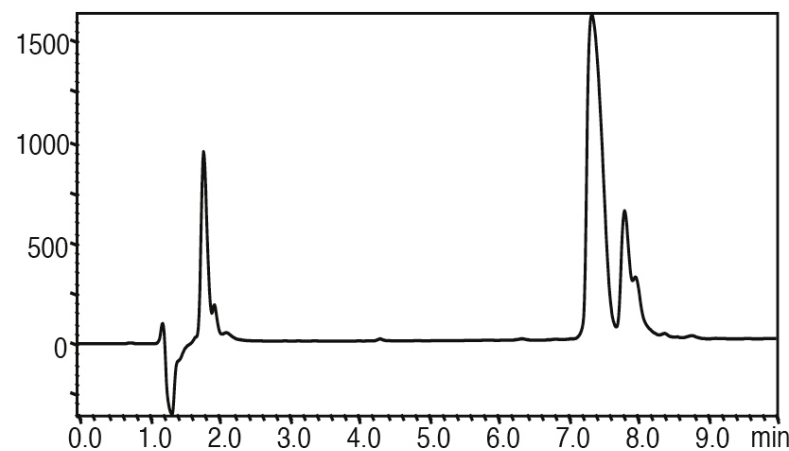
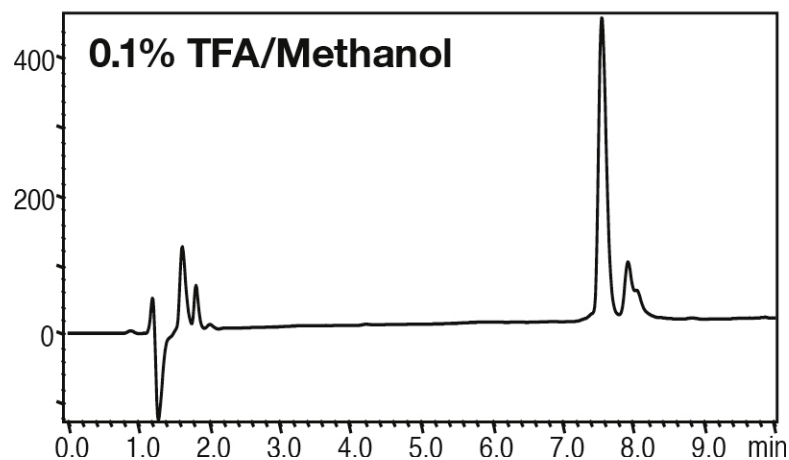
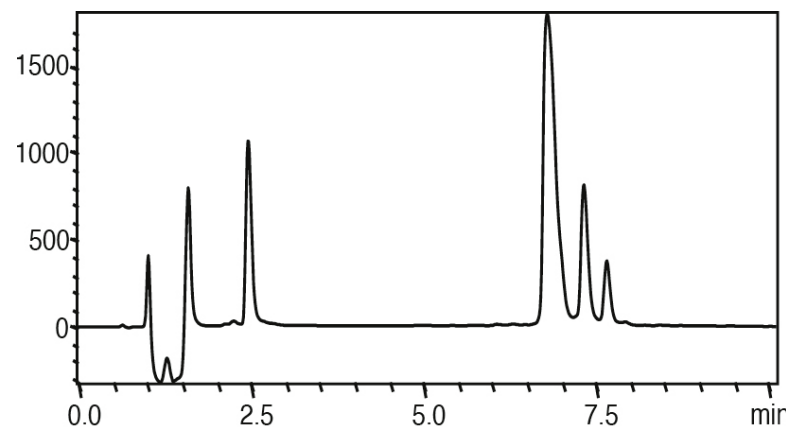
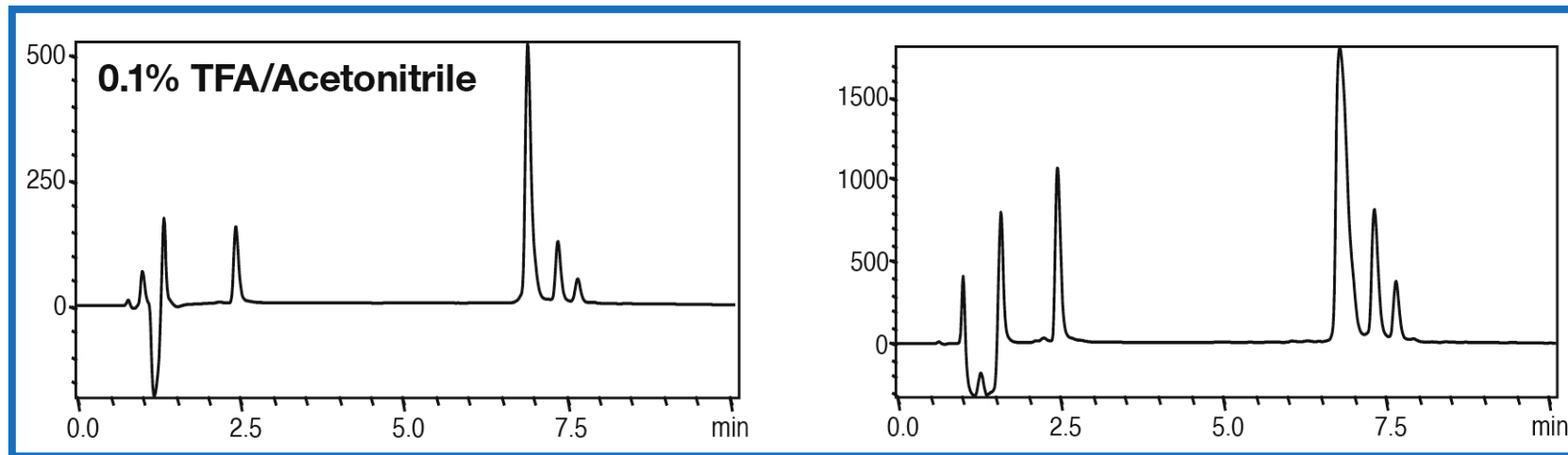
Selection of Organic Solvent for Liraglutide

Loading amount

3 μg

x 8 Loading

24 μg



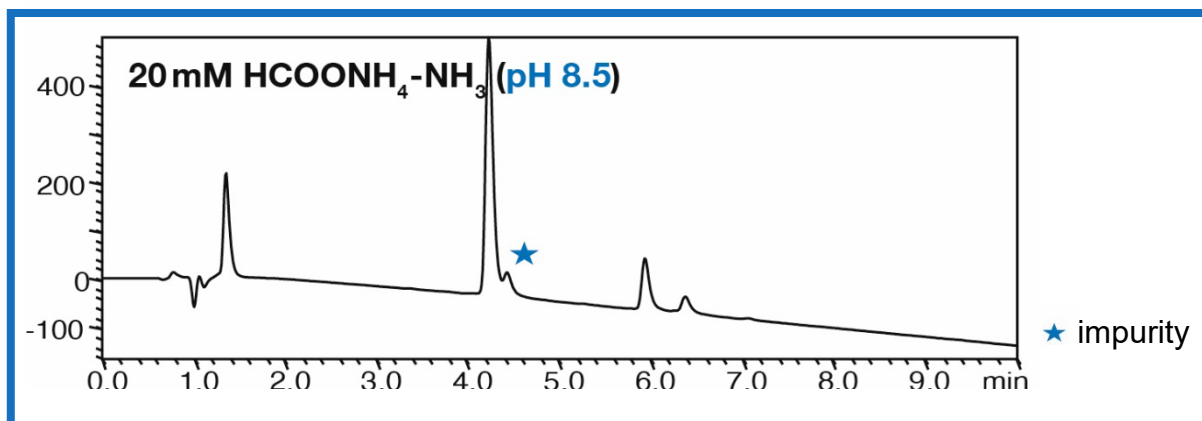
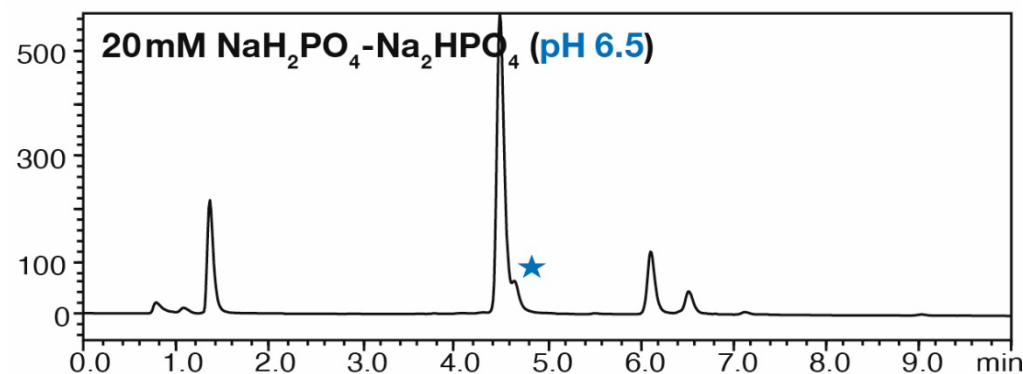
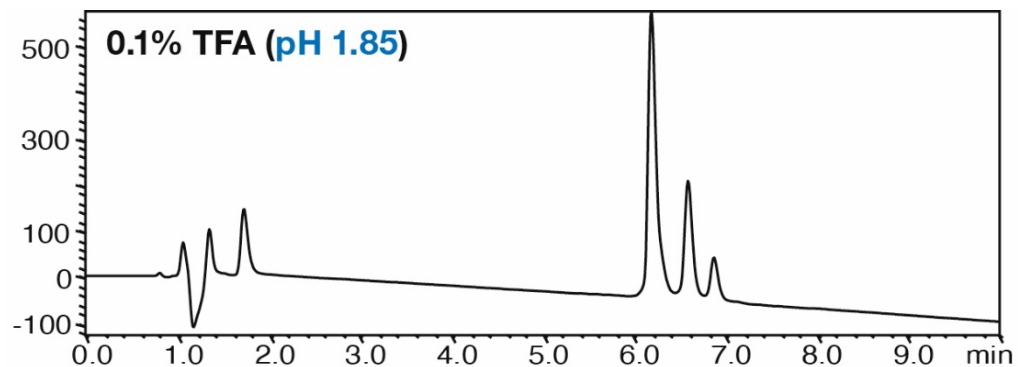
Choice of organic solvent influences the selectivity



Improvement of resolution and loadability

Check the solubility of the peptide in different solvents!

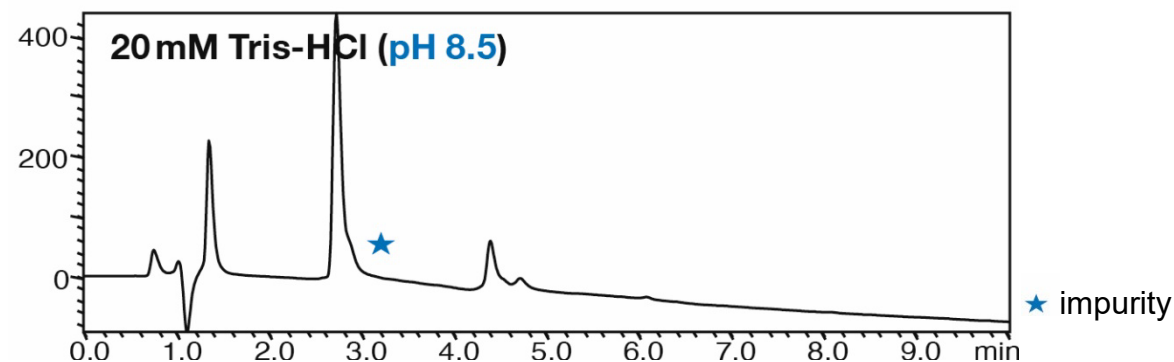
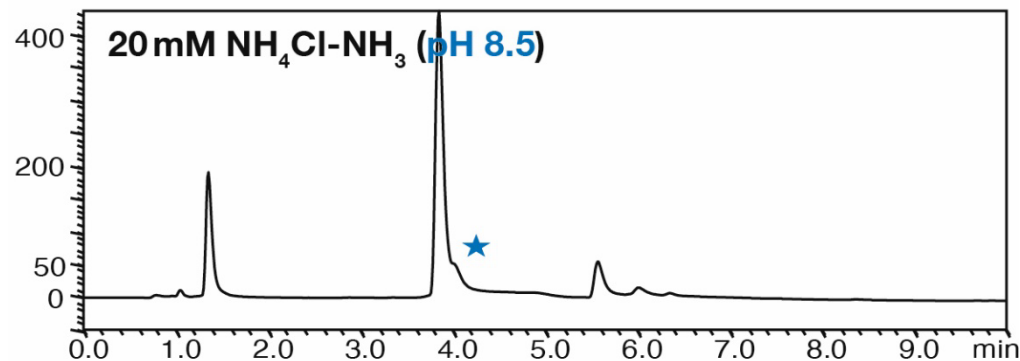
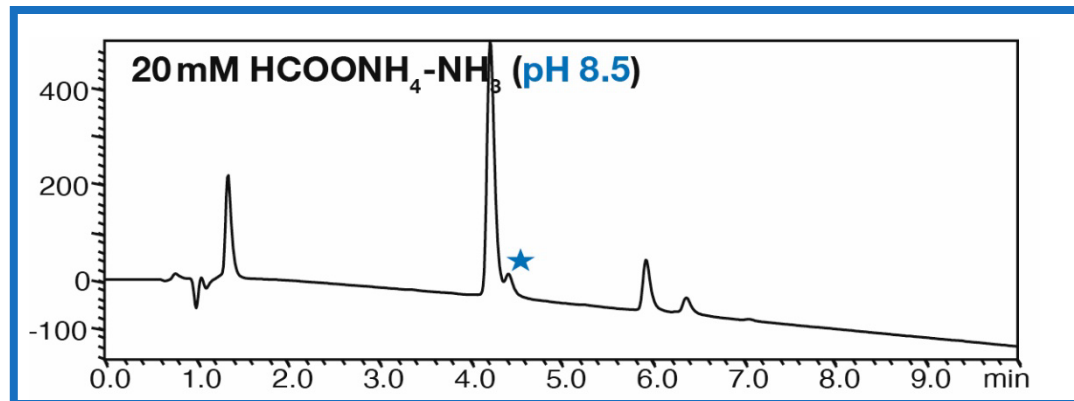
pH Optimisation for Liraglutide



Conditions	Resolution of main peak and impurity peak
20 mM HCOONH ₄ -NH ₃ (pH 8.5)	0.96
20 mM NaH ₂ PO ₄ -Na ₂ HPO ₄ (pH 6.5)	0.50
0.1% TFA (pH 1.85)	-

Maximum flexibility in method development requires highly stable stationary phases!

Buffer Type Selection for Liraglutide



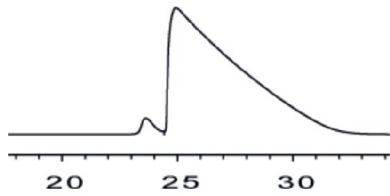
Conditions	Resolution of main peak and impurity peak
20 mM HCOONH ₄ -NH ₃ (pH 8.5)	0.96
20 mM NH ₄ Cl-NH ₃ (pH 8.5)	-
20 mM Tris-HCl (pH 8.5)	-

Buffer salts have a strong influence on the peptide separation
→ Screening is recommended

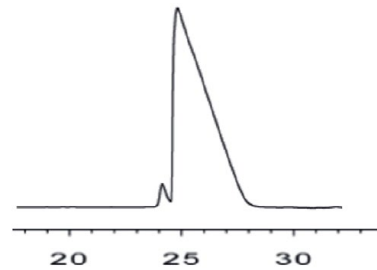
Gradient Optimisation for Liraglutide

Influence on peak shape

Isocratic elution vs. gradient elution

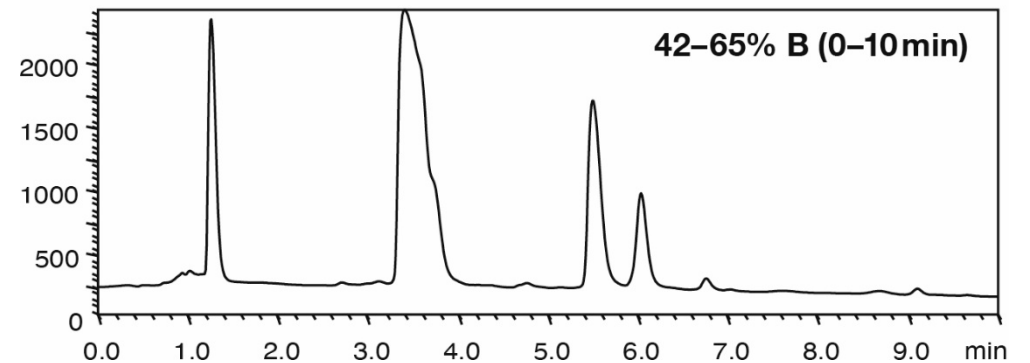
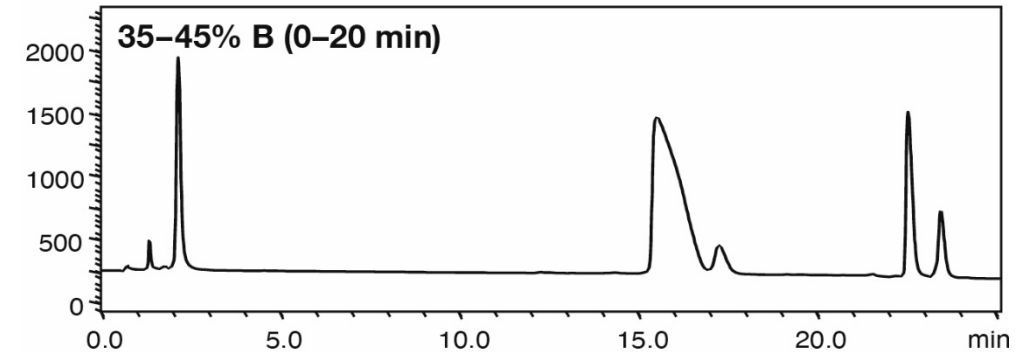
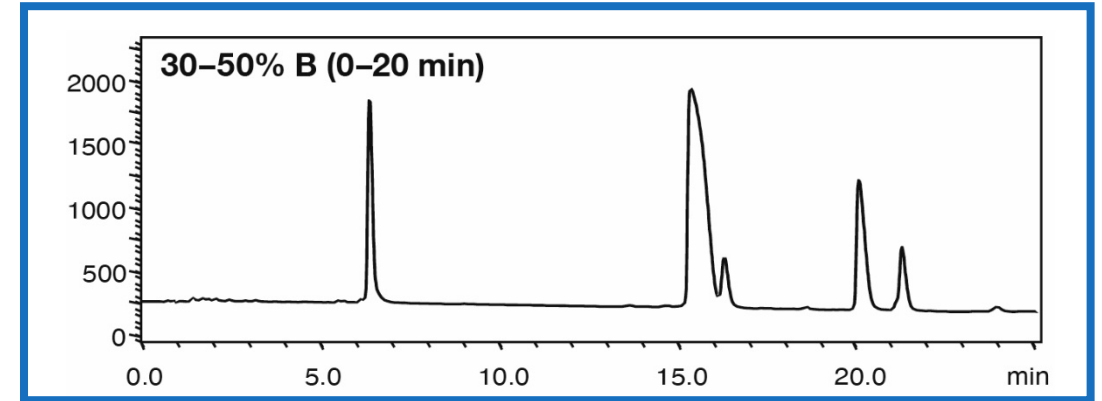


isocratic



gradient

The slope of the gradient has to be adapted to give an optimal balance of resolution and productivity



Conclusion

Development of a purification method is performed in three steps:

1. **Method development at analytical scale**
2. **Loadability studies at analytical scale**
3. **Linear scale-up to preparative process**

Practical considerations for the purification of **Liraglutide**:

- ✓ **pH stable hybrid silica** as base particle for maximum flexibility in method development
- ✓ **Alkaline pH** for highest resolution
- ✓ **Acetonitrile** as organic modifier
- ✓ **Optimised gradient** to increase the loadability