

Reversed Phase Method Development



Step 1

Characteristics

Applicational characteristics

- HPLC/UHPLC-method?
- Which detector? LC/MS?
- Isocratic/gradient?

Analyte characteristics

- Hydrophobicity, polarity, ionicity
- Structure/molecular weight
- Stability
- How do the analytes differ?
- Matrix

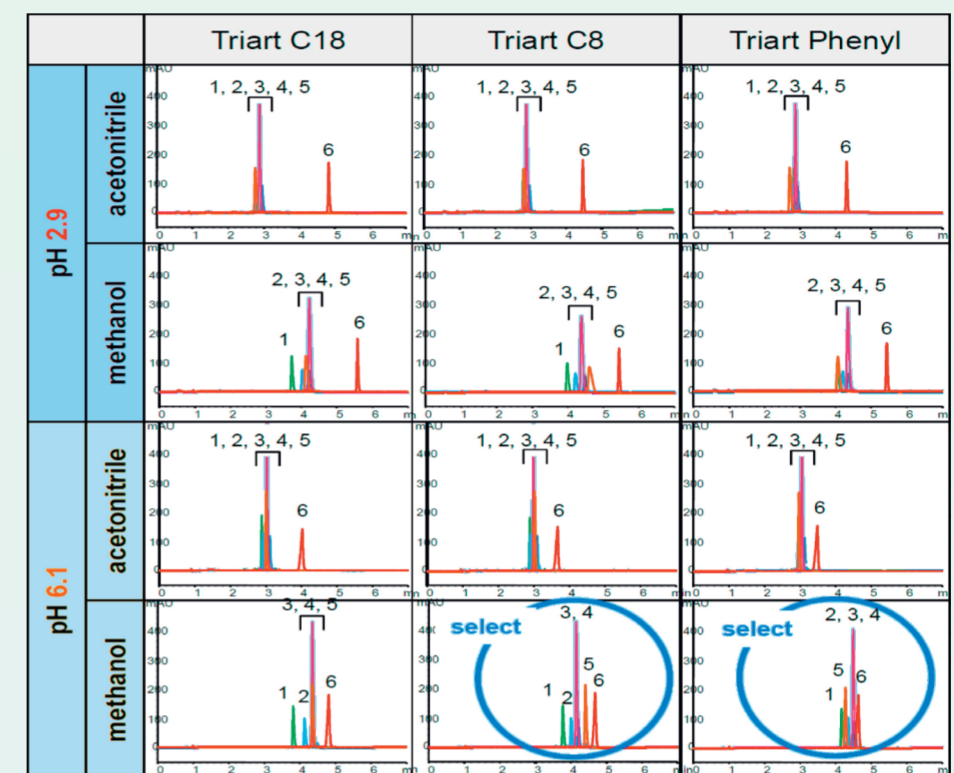
Step 2

Screening

Typical screening conditions:

- Steep gradient
- Mostly short column e.g. 50 mm
- ID depends on compound, pressure

Choose the most promising conditions



Example 6 pigments on YMC-Triart columns
 Column: 50 x 2.0 mm ID
 Gradient: 5-90%B (0-5 min), 90%B (5-7 min), 5%B (7-12 min)
 Flow rate: 0.2 mL/min
 Temperature: 40°C
 Detection: UV at 254 nm

1. Acid green 16
2. Acid blue 1
3. Acid red 52
4. Acid blue 3
5. Methyl orange
6. Methyl red

Columns

Screening kit

1. YMC-Triart C18
2. YMC-Triart C18 ExRS
3. YMC-Triart C8
4. YMC-Triart Phenyl
5. YMC-Triart PFP

X

Mobile phase

Solvent

- 1 Acetonitrile
 - 2 Methanol
- Gradient
5-90% solvent

X

pH (solution)

- I. Acidic
- II. Neutral
- III. Basic

Step 3

Optimisation

Adjustment options

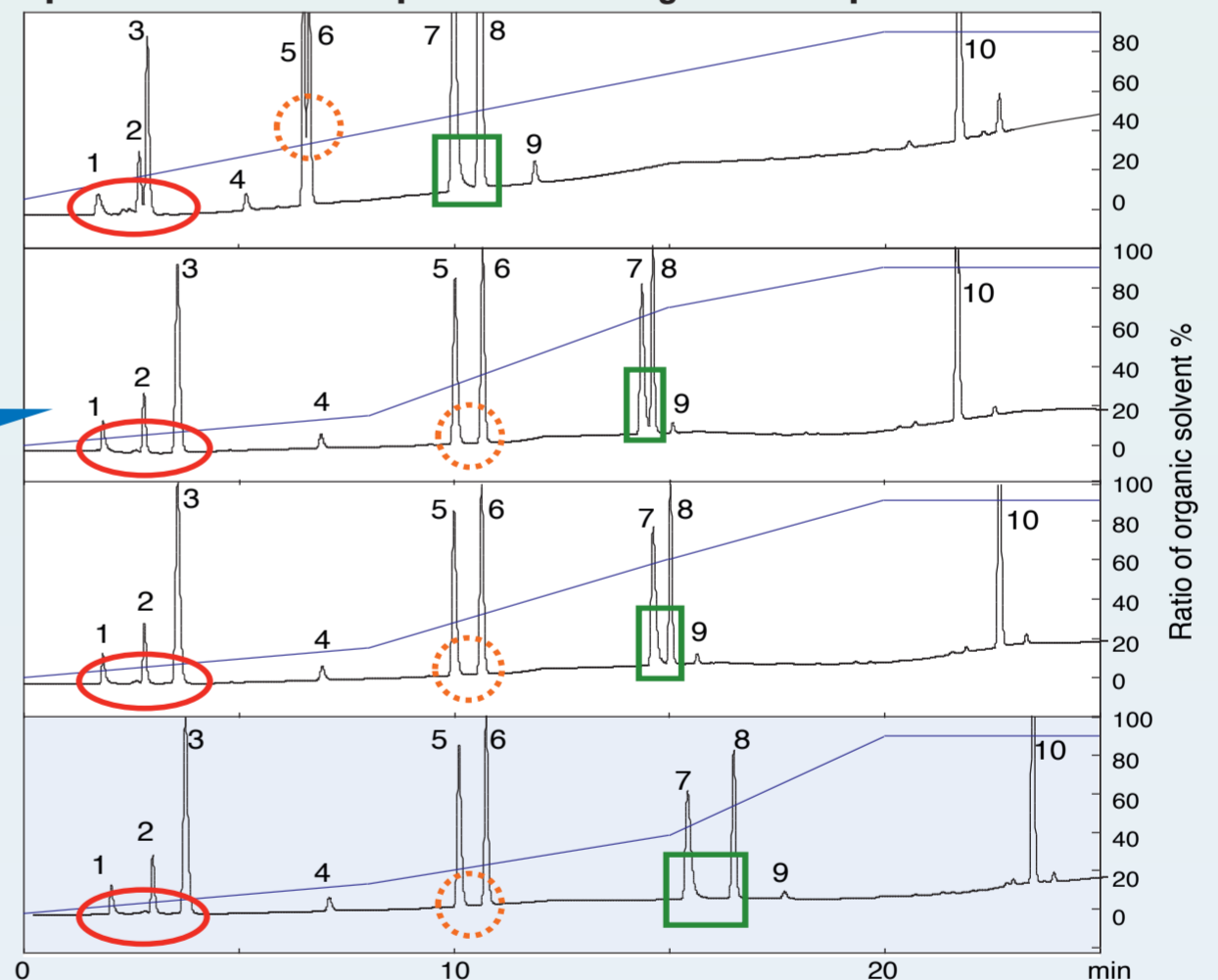
- Mobile phase ratio
- Gradient slope/isocratic
- Precision tuning of temperature
- Column dimensions
- Particle size
- Add org. modifier
- pH (additives)

5-90%B (0-20 min)

Separated by decreasing in gradient slope

0-15%B (0-8 min)
15-40%B (8-15 min)
40-90%B (15-20 min)

Optimisation of mobile phase ratio and gradient slope



Example cold medicine on HydroSphere C18
 Column: HydroSphere C18 (5 µm, 120 Å) 150 x 4.6 mm ID
 Eluent: A) 20 mM phosphate buffer (pH 2.5)
 B) methanol
 Flow rate: 1.0 mL/min
 Temperature: 37°C
 Detection: UV at 210 nm (0-15 min), 235 nm (15-25 min)

1. Thiamine hydrochloride
2. Unknown
3. L-Ascorbic acid
4. Maleic acid
5. di-Methylophedrine hydrochloride
6. Dihydrocodeine phosphate
7. Saccharin sodium
8. Caffeine
9. Chlorpheniramine
10. Ibuprofen

Buffer Selection

Buffers should be chosen as following: **pH 2 values under pKa** of the analyte for acidic compounds and for **basic** compounds **2 values above the pKa**. For LC/MS methods it is recommended to use buffer concentrations <15 mM.

Buffer	pKa	Buffer range [pH]	Standard Concentration	LC/MS compatibility
Trifluoroacetic acid (TFA)	<1.0	-	0.01-0.1%	✓
Phosphoric acid	2.1	-	0.01-0.1%	✗
Ammonium dihydrogen phosphate (Na ⁺ , K ⁺ salt)	2.1	1.1-3.1	5-50 mM (<20 mM recommended)	✗
Formic acid	3.7	-	0.1-1.0%	✓
Ammonium formate (Na ⁺ , K ⁺ salt)*	3.7	2.7-4.7	5-50 mM	NH ₄ ⁺ salt: ✓ [Na ⁺ , K ⁺ salt: ✗]
Acetic acid	4.8	-	0.5-5.0%	✓
Ammonium acetate (Na ⁺ , K ⁺ salt)*	4.8	3.8-5.8	5-50 mM	NH ₄ ⁺ salt: ✓ [Na ⁺ , K ⁺ salt: ✗]
Ammonium hydrogen phosphate (Na ⁺ , K ⁺ salt)	7.2	6.2-8.2	5-50 mM (<20 mM recommended)	✗
Triethylamine acetic acid (TEAA)	-	4.6-6, 10-11	<20 mM	✓
Ammonium formate, ammonium acetate**	9.2	8.2-10.2	<20 mM	✓
Sodium phosphate, potassium phosphate	12.3	11.3-13.3	<10 mM	✗
Ammonium bicarbonate**	-	8.5-10.5	<10 mM	✓

*adjusted with acid **adjusted with ammonia

YMC-Triart Phase Specifications

	C18	C18 ExRS	Bio C18	C8	Bio C4	Phenyl	PFP
Base	organic/inorganic silica						
Stationary phase	C18 (USP L1)	C18 (USP L1)	C18 (USP L1)	C8 (USP L7)	C4 (USP L26)	Phenyl (USP L11)	Penta-fluorophenyl (USP L43)
Particle size	1.9, 3 and 5 µm						
Pore size	12 nm	8 nm	30 nm	12 nm	30 nm	12 nm	12 nm
Carbon content	20%	25%	-	17%	-	17%	15%
Endcapping	multi-stage	multi-stage	multi-stage	multi-stage	multi-stage	multi-stage	none
pH range	1-12	1-12	1-12	1-12	1-10	1-10	1-8
Temperature range	pH < 7: 90°C pH > 7: 50°C	pH < 7: 90°C pH > 7: 50°C	pH < 7: 90°C pH > 7: 50°C	pH < 7: 90°C pH > 7: 50°C	pH < 7: 90°C pH > 7: 50°C	50°C	50°C
100% aqueous eluents	✓	✗	✓	✗	✓	✓	✓